

## Original Research Article

<https://doi.org/10.20546/ijcmas.2020.902.049>

## Cultivation of *Spirulina* using Low-Cost Organic Medium and Preparation of Phycocyanin Based Ice Creams

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### ABSTRACT

*Spirulina platensis* is a filamentous Cyanobacterium with a high content of protein and essential vitamins that are commercially produced for feed and food supplementation. The present study was carried out to utilize the fermented oil cake filtrate as organic nutrient supplementation for *Spirulina platensis* cultivation. Commonly, the cultivation of *S. platensis* is done with conventional Zarrouk media that requires a high quality of NaHCO<sub>3</sub>. To reduce the cost of the medium different concentration of baking soda, sea salt, along with fermented oil cake filtrate were used as media for the cultivation of *Spirulina platensis*. Since there is a greater demand for organically grown *Spirulina* this study was undertaken. The main objective of this work is to cultivate *Arthrospira S. platensis* in edible oil cake incorporated medium. Higher biomass and pigment production concentration were scaled up to form labeled photobioreactor to outdoor trough cultivation. The nutritional content of the organic media was supplemented with the oil cake filtrate. The oil cake filtrate having a rich amount of nitrogen and phosphates will enhance the growth of *Spirulina*. In the present study, the *Spirulina* growth performance was recorded in both traditional and organic media of these the latter recorded better results. The phycocyanin of food-grade purity was incorporated in ice cream from organically cultivated biomass with food-grade and its sensory evaluations were done.

#### Keywords

*Spirulina platensis*, Zarrouk media, oil cakes, organic media phycocyanin, ice creams.

#### Article Info

Accepted:  
05 January 2020  
Available Online:  
10 February 2020

### Introduction

*Spirulina*, a Bluegreen alga having high nutritive value, exists in freshwater, brackish water and seawater habitats. It also can grow in warm temperatures and tropical regions and it is reported to grow abundantly in

alkaline water bodies and salty lakes. *S. platensis* has been used as food for many centuries by different populations and only discovered in recent years. The annual production of the algae is about 10,000 tons which makes it the largest microalgal cultivation industry in the world, (Richmond,

*et al.*, 2000). *Spirulina* is a filamentous cyanobacterium with a high content of protein, essential vitamins that are commercially produced as food and feed supplementation.

The main objective of the present work is to cultivate the *S.platensis* using edible oilcake filtrate incorporated organic medium. Compared to other microalgae the overall production of *Spirulina* has increased from 5000 to 15000 tons /per year.

The production of *Spirulina* increase occurs due to the rich protein (60-70%) composed of essential amino acids, vitamins include B<sub>12</sub>, polyunsaturated fatty acids, pigments and phenolic compounds due to their rich nutrient content has been marketed as a food supplement (Thajuddin and Subramanian 2005). The *S.platensis* have promising bioactive compounds compound viz, phycobiliproteins, phenolic compounds, organic acids, beta carotene, polyunsaturated fatty acids, polysaccharides etc.,(Belay *et al.*, 2002 and Ozdemir *et al.*, 2004)

These bioactive compounds have potential bioactive properties such as reducing the blood cholesterol levels, preventing cancer, stimulating the immune system to reduce the nephrotoxicity of pharmaceuticals. Monchai Dejsungkranont *et al.*, (2013) optimized the algal biomass production experiments statistically designed variables varied viz, light intensity, pH, two different strains of Cyanobacteria, different concentrations of Zarrouk medium, rate of aeration mixed with CO<sub>2</sub> and temperature were studied through the of Taguchi method.

Dinesh Kumar *et al.*, (2015) reported that the organic additive used enhanced the growth of *Spirulina platensis*. Centrifuge supernatant solution of Molasses was used as the organic additive for the *Spirulina* cultivation.

Molasses was used after centrifugation at 5000 rpm for 10 minutes. The supernatant solution was used in different concentrations for enhancing the growth of *Spirulina platensis*. Rajeswari and Deepika, (2017) formulated the organic media for *Spirulina* cultivation by using agro wastes like beetroot, grapes, rice bran, the root of *Casuarina*, etc..

## **Materials and Methods**

### **Sample collection and pre culturing**

*Spirulina platensis* culture MK343101 was obtained from the Department of Biology, Gandhigram Rural Institute-Deemed to be University, Dindigul. The starter inoculums and culture were maintained in the traditional Zarrouk medium. The present study emphasis was on the use of the oil cake filtrate medium, baking soda along with that sea salts for the formulation of organic media for the cultivation of *Spirulina platensis*.

Different concentration of baking soda viz., 20-100%, sea salt and ten ml of the fermented oil cake filtrate has been used for the media formulation. Preliminary work was done to standardize ten ml of the oil cake filtrate was used for 1 L of organic media.

### **Microscopic identification**

The procured cultures were identified using the morphological characters (spiral shape) of blue-green algae (*Spirulina*) identified through the light microscope (Hernando Cortez 1519).

### **Cultivation of *Spirulina***

*Spirulina* was cultured in oil cake filtrate mediums in 1 L Pyrex glass photobioreactor under a controlled condition and Zarrouk medium as control. The cell concentration was analyzed at OD 560 nm Leduy *et al.*,

(1997). The pH of the medium was measured by using a pH meter. *Spirulina platensis* grows at a pH range of 8.9 to 9.5 ± 0.2 as it is maintained at the alkaline point.

Then culture was allowed to grow for 10 days and the biomass was harvested by filtration method, washed with distilled water to remove impurities and neutralize the pH.

### **Estimation of protein content**

The protein content of the harvested *Spirulina* biomass obtained by different organic media protein content was analyzed by Lowry's method (Lowry *et al.*, 1951).

### **Estimation of Beta-carotene**

Beta carotene is one of the major bioactive compounds present in the *Spirulina platensis*. The extraction of the Beta carotene was done through the methodology outlined by a with a slight modification of (Mandelli *et al.*, 2012)

### **Estimation of Phycocyanin**

Algal biomass sample of 0.1 g was mixed with 10ml of citric acid buffer (pH 6.8). This mixture was kept in a deep freezer at -20° C for 24 hours and thawed for every 6 hours respectively. After thawing the mixture was centrifuged at 5000 rpm for 20 minutes.

The resulted supernatant was blue in color it denotes the presence of Phycocyanin pigment present in the biomass. The supernatant was read under a UV visible Spectrophotometer at 620nm. The crude phycocyanin was calculated through the formula (Boussiba and Richmond, 1979).

$$\% \text{ CPC} = \frac{\text{A}_{620} * 10 \text{ ml} * 100}{7.3 * \text{mg sample}}$$

Where 7.3 is the Extinction coefficient of CPC at 620 nm.

### **Purity analysis of phycocyanin**

The purity of the phycocyanin can be calculated through the following formula  
Purity = A<sub>620nm</sub>/A<sub>280nm</sub> (Bennett and Bogorad, 1973).

### **Concentrated the crude phycocyanin**

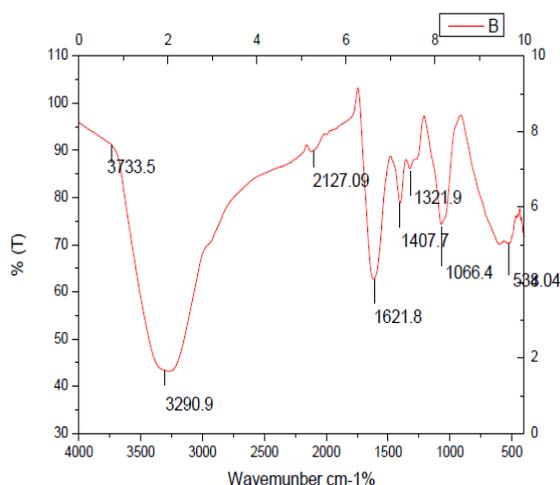
Phycocyanin was concentrated through the rotary vacuum evaporator. The extracted crude phycocyanin was poured into the rotary vacuum chamber at for 40° C at 20 mbar with 5 rpm in 20 minutes to concentrate it. Then the concentrated Phycocyanin was freeze-dried and used for further analysis.

### **The purity of the concentrated phycocyanin**

The purity of the concentrate phycocyanin was analyzed through the UV-VIS spectroscopy at 620 nm and 280nm.(Rito-Palomares *et al.*, 2001) wherein A<sub>620</sub> is the maximum absorbance of C-PC and A<sub>280</sub> is the absorbance of total proteins. The purity ratio meets if above 0.9 indicates it to be a food-grade fit for the food preparation.

### **FT-IR analysis of concentrated Phycocyanin**

Concentrated crude phycocyanin was used for the FT-IR analysis to detect the antimicrobial functional compound through the ATR method. The surface functional group of Phycocyanin extracted from *Spirulina*, were unraveled by FT/IR 4700typeA. The Phycocyanin sample directly poured onto the crystal. The whole crystal-covered and performing quantitative or qualitative analysis. Amin Seyed Yagoubi *et al.*, (2017)reported the characterization and evaluation of physicochemical properties of phycocyanin loaded solid lipid nanoparticles and nanostructured lipid carriers.



**Figure.1** FT-IR Spectral graph of concentrated phycocyanin.

The IR spectral peaks indicate antimicrobial products of the phycocyanin bands at 1321 cm-1, 1407 cm-1 (C-O-C), 1621 cm-1 (CO of amide), 2127 cm-1 (C = C) and broadband 3290 cm-1 (of OH and NH), Mostafa *et al* .,(2014) observed the antimicrobial substance from the *S.platensis*, in that study they found the spectroscopic analysis of the purified antimicrobial product IR spectrum showed bands at 1269 cm-1, 1414 cm-1 (C-O-C),

1643cm-1 (CO of amide),1563 cm-1 (C = C) and broadband 3441 cm-1 (of OH and NH).

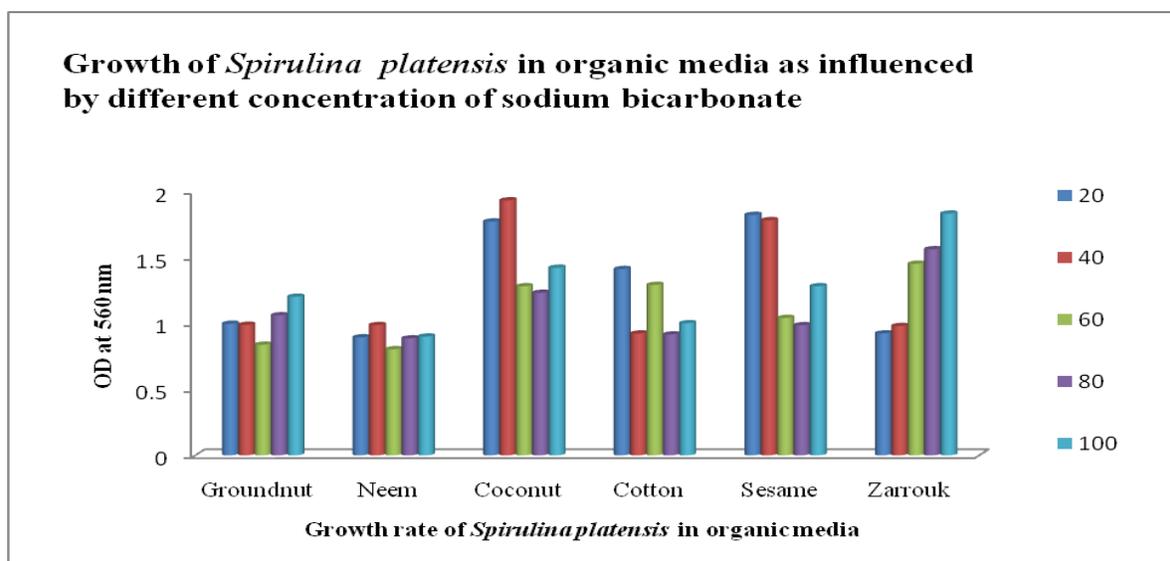
### Microbial analysis of Phycocyanin

The microbial content of the crude phycocyanin was analyzed standard plating method was followed. The standard plate count method was followed. One ml of concentrated phycocyanin was taken for the microbial analysis.

### Ice cream preparation

Ice creams were prepared by the standard method, during the freezing time the concentrated phycocyanin added into 50gm of ice cream at 2ml, 5ml, 8ml, 10ml.

Then mixed and frozen leave the ice cream in the freezer for a final freezer, until solid. The main sensory characteristics were evaluated as mentioned outlined by (Abd El Baky *et al.*, 2015)



**Figure.2** Growth of *Spirulina platensis* in organic media as influenced by different concentration of sodium bicarbonate

After 10 days of culture, the absorbance of all-organic medium and control OD reached above 0.8. The coconut filtrate with 40% sodium bicarbonate added medium reveals the highest growth OD as (1.93) and the lowest growth rate was observed in neem filtrate with 60 % sodium bicarbonate medium growth OD (0.803).

Mahavir Joshi *et al.*, 2014 demonstrated Zarrouk’s media Supplemented along with rainwater, cows urine, well water cheese whey the maximum growth rate was observed at the fifteenth day of all media the maximum OD (560 nm ) at 15 the day such as 1.76, 1.53, 1.75, 1.44. (Fig .1) The biomass content of the

all-organic medium along with control (Zarrouk) were as follows expressed as g/100 ml among the five organic medium as 20% groundnut oil cake filtrate medium showed 0.81 ±0.197 gm/100 ml, 40% sesame medium shows the 1.14±0.325 60% coconut medium 1.455±0.473 and 80% groundnut medium 1.405±0.700 and the 100 % coconut oil cake medium 1.395±0.304.

Jain and Singh (2013), they formulated a low-cost medium using cow dung ash for the biomass production they reported that *Spirulina platensis* is capable to grow in various kinds of culture media obtained 1.212 g/L dry biomass.

**Table.1** Optimization of biomass production as influence by different oil cake filtrate media with different concentrations of sodium bicarbonate

S.no	Substrates oil cake	20% NaHCO <sub>3</sub> (gm /100ml)	40% NaHCO <sub>3</sub> (gm/100ml)	60% NaHCO <sub>3</sub> (gm/100ml)	80% NaHCO <sub>3</sub> (gm/100ml)	100% NaHCO <sub>3</sub> (gm/100ml)
1	Neem	0.7 ±0.197	0.45 ±0.183	0.71 ±0.339	0.915±0.007	1.185±0.714
2	Coconut	0.3±0.042	0.49 ±0.113	1.455±0.473	1.09 ± 0.395	1.395±0.304
3	Groundnut	0.81±0.197	1.05 ±0.197	0.915±0.572	1.405±0.700	1.18 ± 0.480
4	Cotton	0.23±0.02	0.58 ±0.240	0.975 ± 0.26	1.165±0.261	1.16 ± 0.353
5	Sesame	0.63±0.183	1.14± 0.325	0.965±0.275	1.235±0.148	0.885±0.388
6	Zarrouk	0.43±0.13	0.67 ± 0.14	0.98 ± 0.53	1.45± 0.10	1.66 ± 0.14

(gm/100 ml wet biomass)

**Table.2** Determination of protein content in different oil cake filtrate media

S.no	Substrates(Oil cake)	20% NaHCo3 g/100 gm	40% NaHCo3 g/100 gm	60% NaHCo3 g/100 gm	80% NaHCo3 g/100 gm	100% NaHCo3 g/100 gm
1	Neem	37.83±0.9	56.31±0.75	40.88±0.54	37.16±0.54	35.08±0.61
2	Coconut	40.23±0.52	42.68±0.36	35.11±0.09	30.28±0.08	29.39±0.61
3	Groundnut	33.43±0.79	57.23±0.54	42.68±0.54	33.28±0.09	30.35±1.05
4	Cotton	35.82±0.43	53.23±0.46	39.04±0.04	41.6±0.26	34.37±0.8
5	Sesame	42.15±0.69	54.06±0.12	40.06±0.59	42.44 ±0.62	32.64±0.66
6	Zarrouk	56±0.64	61.39±0.54	61.75±0.43	60.67±0.54	64.42± 0.46

Aruna *et al.*, 2008 reported 10 g of groundnut cake in 1 L, 15 g of neem cake, 15 g of press mud along with baking soda and one gram of sea salt used media enhance the growth of *Spirulina* and gave better results such as maximum growth rate were reported  $1.0 \pm 0.1$  and chlorophyll  $0.09 \pm 0.07$   $\mu\text{g/ml}$  and protein  $45 \pm 3\%$  and neem cake media  $0.9 \pm 0.1$  chlorophyll  $0.09 \pm 0.07$   $\mu\text{g/ml}$  and protein  $45 \pm 3\%$ .

Organic media from agro-waste like skin peel from beetroot (*Beta vulgaris*), leaves grape (*Vitis vinifera*), and extracts from rice bran and rice husk and root from Casuarina (*Casuarina equisetifol*) skin peel of beetroot grape leaves an extract of rice and improve the growth of *Spirulina* Rajeshwari and Deepika (2017). Table 1 provides the yield of

biomass from the various concentration of sodium bicarbonate and oil cake filtrated treatment.

The table results revealed that the different concentrations of sodium bicarbonate along with the organic oil cake filtrate as the media for the growth of *Spirulina platensis* for this study, it has to influence the growth of *Spirulina* at different levels.

Compared to the Zarrouk media lowest concentration of sodium bicarbonate and groundnut oil cake filtrate medium gave the highest biomass production with other media. Table 2 shows the protein content of the various concentration of sodium bicarbonate and oil cake filtrated treatment.

**Table.3** Determination of Beta-carotene in different oil cake filtrate media

S.no	Substrates (Oil cake)	20% ( $\mu\text{g/gm}$ )	40% ( $\mu\text{g/gm}$ )	60% ( $\mu\text{g/gm}$ )	80% ( $\mu\text{g/gm}$ )	100% ( $\mu\text{g/gm}$ )
1	Neem	1065 $\pm$ 0.37	2013 $\pm$ 0.79	1756 $\pm$ 0.76	1137 $\pm$ 1.39	541 $\pm$ 0.59
2	Coconut	1038 $\pm$ 1.88	134 $\pm$ 1.54	1146 $\pm$ 0.78	584 $\pm$ 0.979	494 $\pm$ 0.54
3	Groundnut	1054 $\pm$ 1.81	2184 $\pm$ 0.65	1877 $\pm$ 2.26	1141 $\pm$ 1.39	686 $\pm$ 1.9
4	Cotton	1164 $\pm$ 0.32	2008 $\pm$ 0.57	1386 $\pm$ 0.49	698 $\pm$ 0.99	653 $\pm$ 0.34
5	Sesame	1145 $\pm$ 1.5	1457 $\pm$ 0.72	1489 $\pm$ 0.33	998 $\pm$ 0.65	766 $\pm$ 1.7
6	Zaarrouk	1040 $\pm$ 0.59	1421 $\pm$ 0.32	1488 $\pm$ 0.45	1546 $\pm$ 0.74	1817 $\pm$ 0.34

It represents beta-carotene of the various concentration of sodium bicarbonate and oil cake filtrated treatment.

**Table.4** Sensory evaluation of phycocyanin based ice cream

Sensory characters	Control without phycocyanin	Phycocyanin with different concentration
Colour	9.7 $\pm$ 1.02	8.7 $\pm$ 1.18
Taste	10 $\pm$ 0.14	9.4 $\pm$ 1.23
Texture	10 $\pm$ 0.43	8.9 $\pm$ 1.14
Aroma	9.9 $\pm$ 0.02	8.7 $\pm$ 1.63
Overall acceptability	9.9 $\pm$ 0.1	8.7 $\pm$ 1.20

(Mean  $\pm$ Standard deviation n=10)

### Preparation of Ice-cream supplemented with concentrated phycocyanin

The Food Function Products (FFP) were

prepared by using the standard ice cream preparation method. The food-grade pc purity (0.9) level of phycocyanin was incorporated into ice cream at different concentration levels (w/v) concentrated phycocyanin was

added as an active ingredient at 2 to 10 ml to the ice cream and commercial ice cream kept as a standard control.

A control ice cream without any phycocyanin was also prepared. All the ice cream were prepared in cold condition. After preparation stored in the freezing condition, ice creams were protected from light and temperature.

The ice creams were prepared with control ice cream. The preparation with maximum phycocyanin incorporation (10ml/ 50gm) in terms of color, texture, taste of the phycocyanin. These results indicate the positive effect of the phycocyanin.

These results revealed that ice cream prepared with higher phycocyanin concentration added ice cream is most likely to the panelist. And the taste leads to a good appreciation. These results revealed that phycocyanin based ice creams are the most favorite and very attractive when we prepared with the addition of phycocyanin.

The present study, investigated the comparative growth rate of *Spirulina* in Zarrouk's medium supplemented with different organic oil cake substrates. *Spirulina* cultivation was evaluated by lab-scale cultivation by using different organic based substrates as growth supplements.

*Spirulina* grows well in all the four substrates; the best growth, protein and pigment content were obtained in neem, groundnut and coconut filtrate supplemented medium. The experimental setup indicates that when *Spirulina* cultivated under varying concentrations of substrate supplementation, yield better growth and could be optimized as a low-cost media for the mass cultivation of *Spirulina*.

That organically cultivated product exhibits

the functional food preparations. This present study extracts the phycocyanin from the organically cultivated *Spirulina platensis*, possesses numerous bioactivities, has been used for the phycocyanin based ice cream preparation, the sensory evaluation of Food Function Products of ice creams are on the bar with the commercial ice creams.

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**How to cite this article:**

Sukanya. A, R. Meena and David Ravindran. A. 2020. Cultivation of *Spirulina* using Low-Cost Organic Medium and Preparation of Phycocyanin Based Ice Creams. *Int.J.Curr.Microbiol.App.Sci.* 9(02): 392-399. doi: <https://doi.org/10.20546/ijcmas.2020.902.049>